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GETHI

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Ilustre Colegio Oficial de Médicos de Madrid. Aula Jiménez Díaz. Madrid

NTRK: ¿Qué alteraciones presentan y en qué tumores?

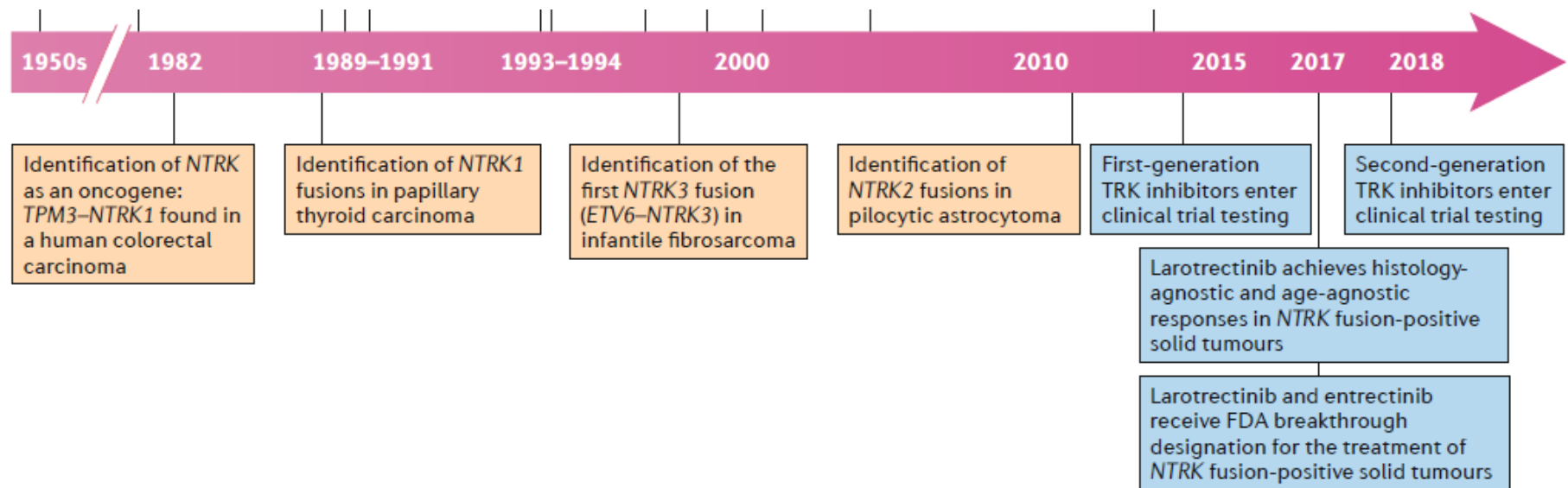


Hospital Universitario
12 de Octubre

Dra. Lara Iglesias

Oncología Médica

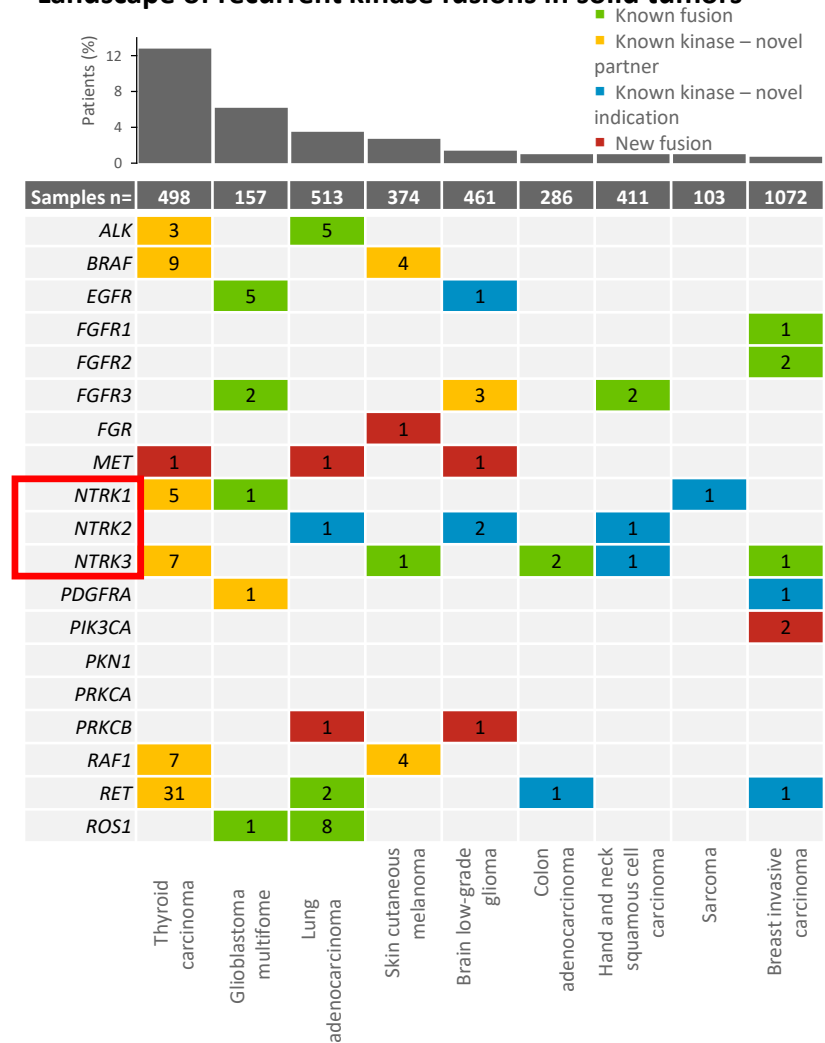
HISTORY OF ONCOGENIC *NTRK* GENE FUSIONS



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- Associated with a diverse range of solid tumors and hematologic malignancies^{1,2}
- Neurotrophic tyrosine receptor kinase (*NTRK*) gene fusions are associated with many human cancers
 - Associated with ≥19 tumor types^{2,3}
 - Implicated in up to 1% of all solid tumors⁴

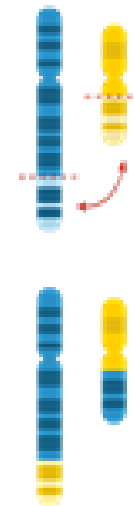
Landscape of recurrent kinase fusions in solid tumors⁵



- Khotskaya YB, et al. *Pharmacol Ther.* 2017;173:58-66
- Vaishnavi A, et al. *Cancer Discov.* 2015;5:25-34.
- Amatu A et al. *ESMO Open.* 2016;1:e000023.
- Drilon A, et al. *N Engl J Med.* 2018;378:731-739.
- Stransky N, et al. *Nat Commun.* 2014;5:846.

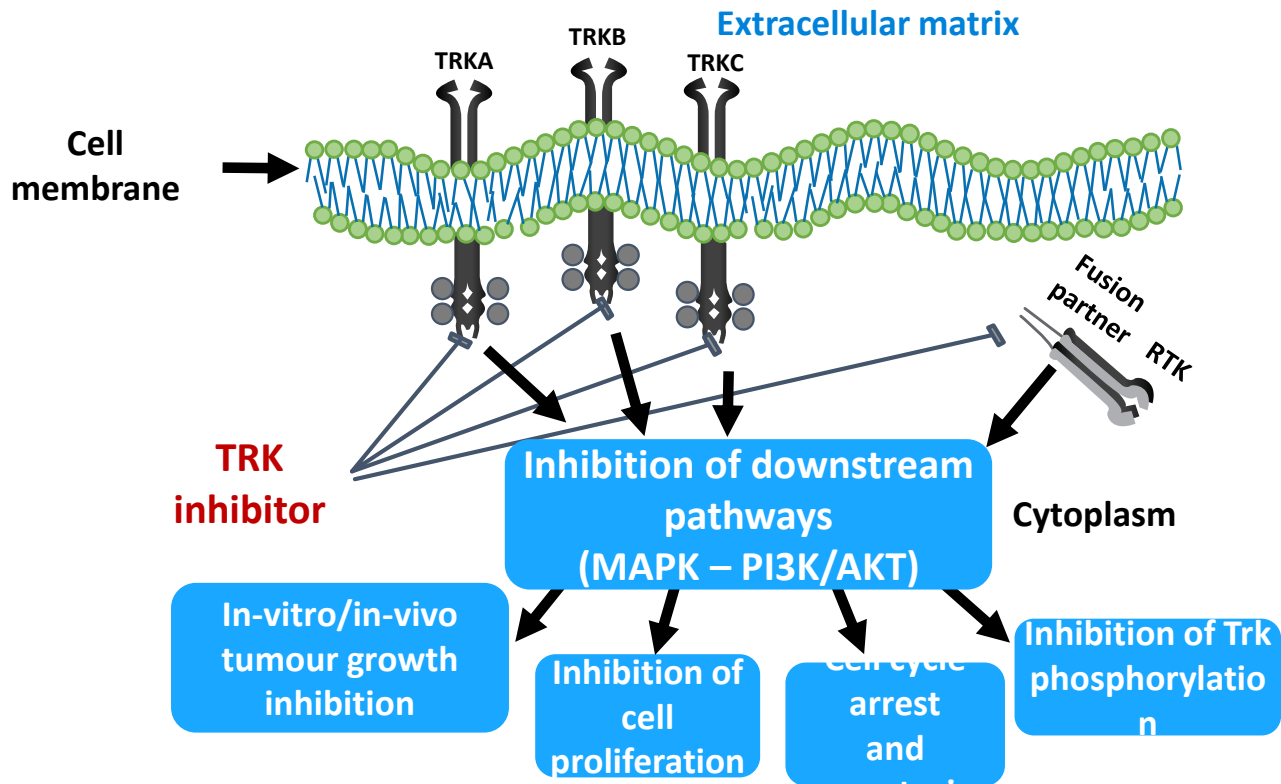
NTRK: Rearrangements/ fusions

- Neurotrophic tyrosine receptor kinase (*NTRK*) gene fusions :
 - involving either *NTRK1*, *NTRK2* or *NTRK3*
 - (encoding the neurotrophin receptors
 - TRKA , TRKB and TRKC, respectively)



2. Vaishnavi A, et al. *Cancer Discov.* 2015;5:25-34.
3. Amatu A, et al. *ESMO Open.* 2016;1:e000023.
4. Drilon A, et al. *N Engl J Med.* 2016;378:731-739.
5. Stransky N, et al. *Nat Commun.* 2014;5:846.

Tyrosine kinase domain of TrkA, TrkB, TrkC receptors¹



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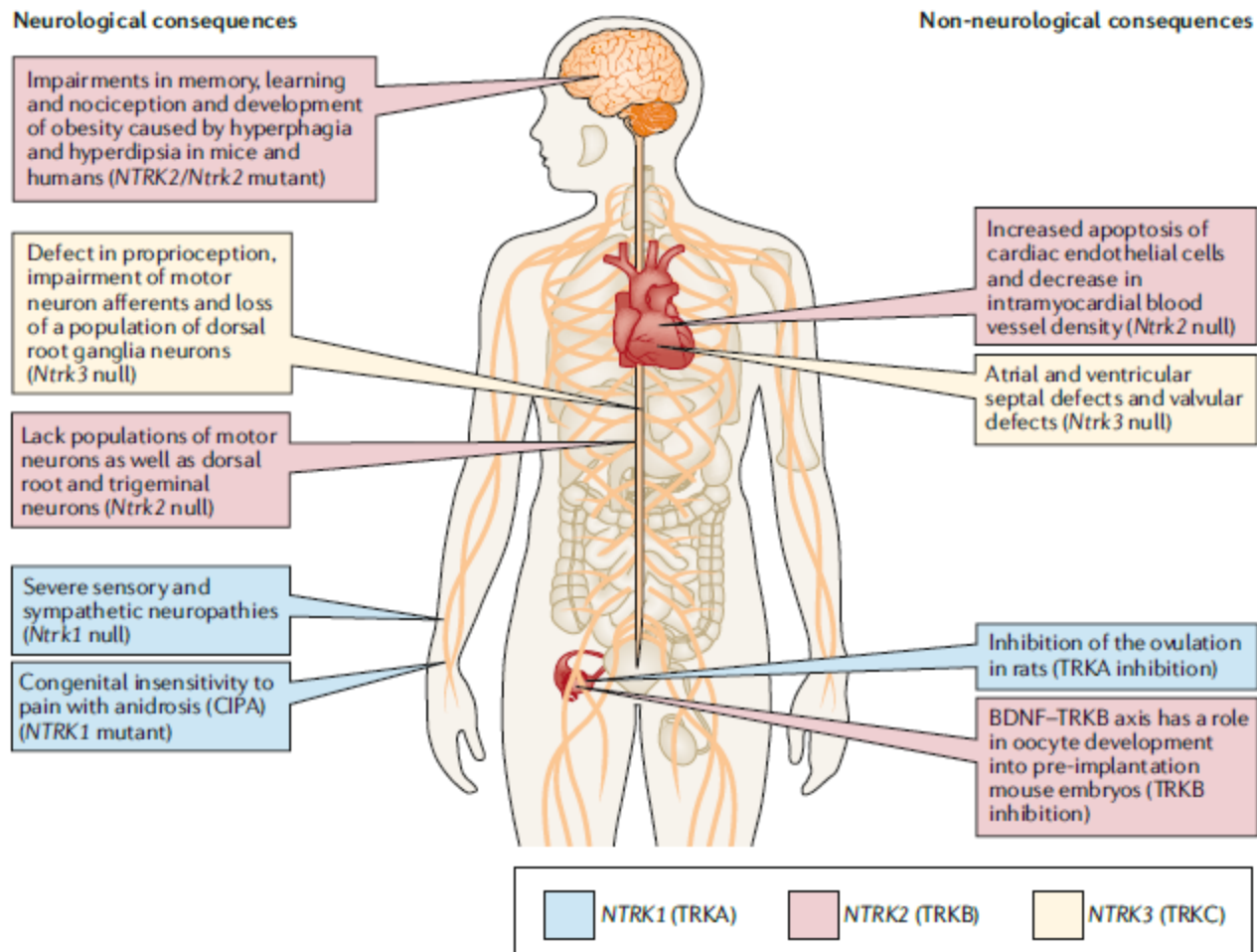
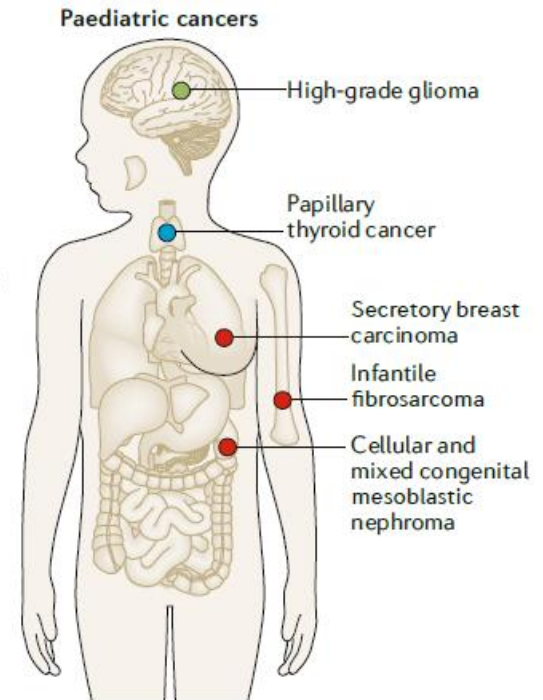
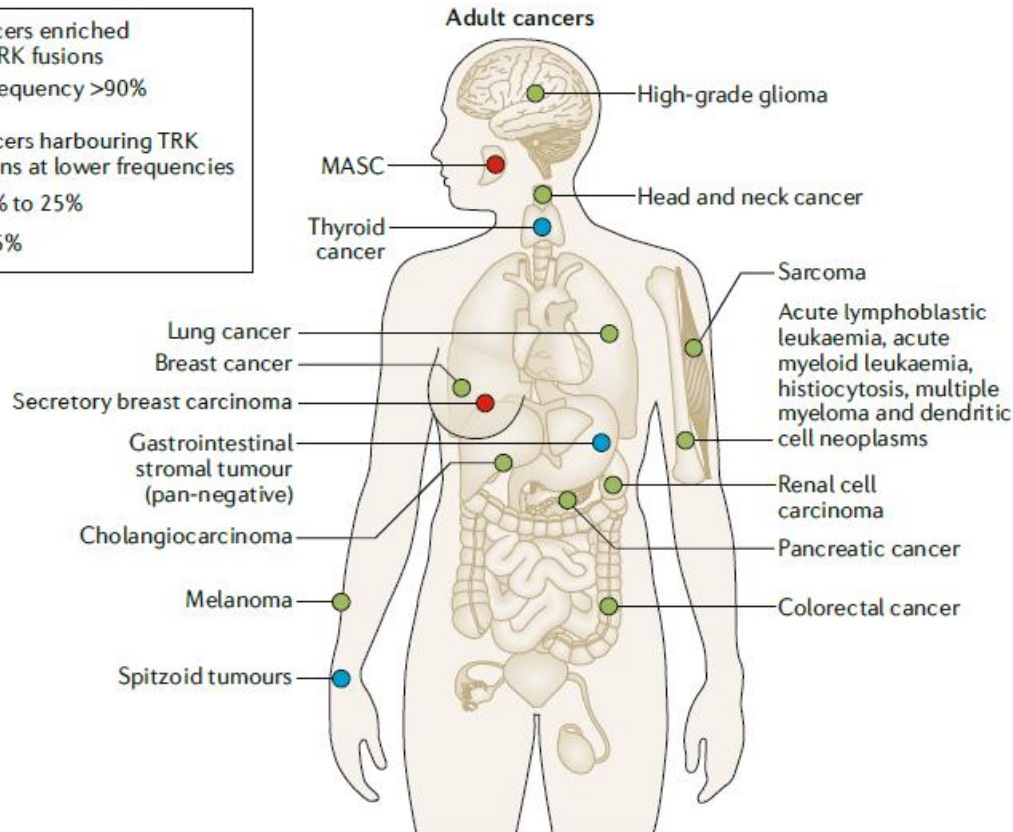


Fig. 6 | Consequences of loss, decreased activity or inhibition of TRK. Genetic or pharmacological disruption of TRK

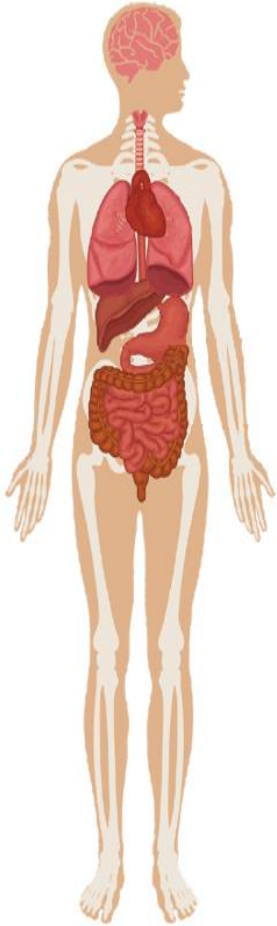
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Cancers enriched for TRK fusions
● Frequency >90%

Cancers harbouring TRK fusions at lower frequencies
● 5% to 25%
● <5%



Adult



Brain cancers (0.4–3.1%)¹



Salivary (MASC; 90–100%)¹



Secretory breast cancer (92%)^{2*}



Pancreatic cancer (<1%)^{3,4}



Cholangiocarcinoma (3.6%)¹



Thyroid cancer (1.5–14.5%)¹



Lung cancer (0.2–3.3%)¹



GIST (3.2%)⁵



Colon cancer (1.5%)¹



Melanoma (0.3%)^{1,6}



Sarcomas (1%)^{6*}

Pediatric



Brain cancers (~10%)⁷



Spitzoid neoplasms (16.4%)⁸



Gliomas (7.1%)¹



Thyroid (9.4–25.9%)^{9,10}



Infantile fibrosarcoma (91–100%)¹¹



Congenital nephroma (83%)¹



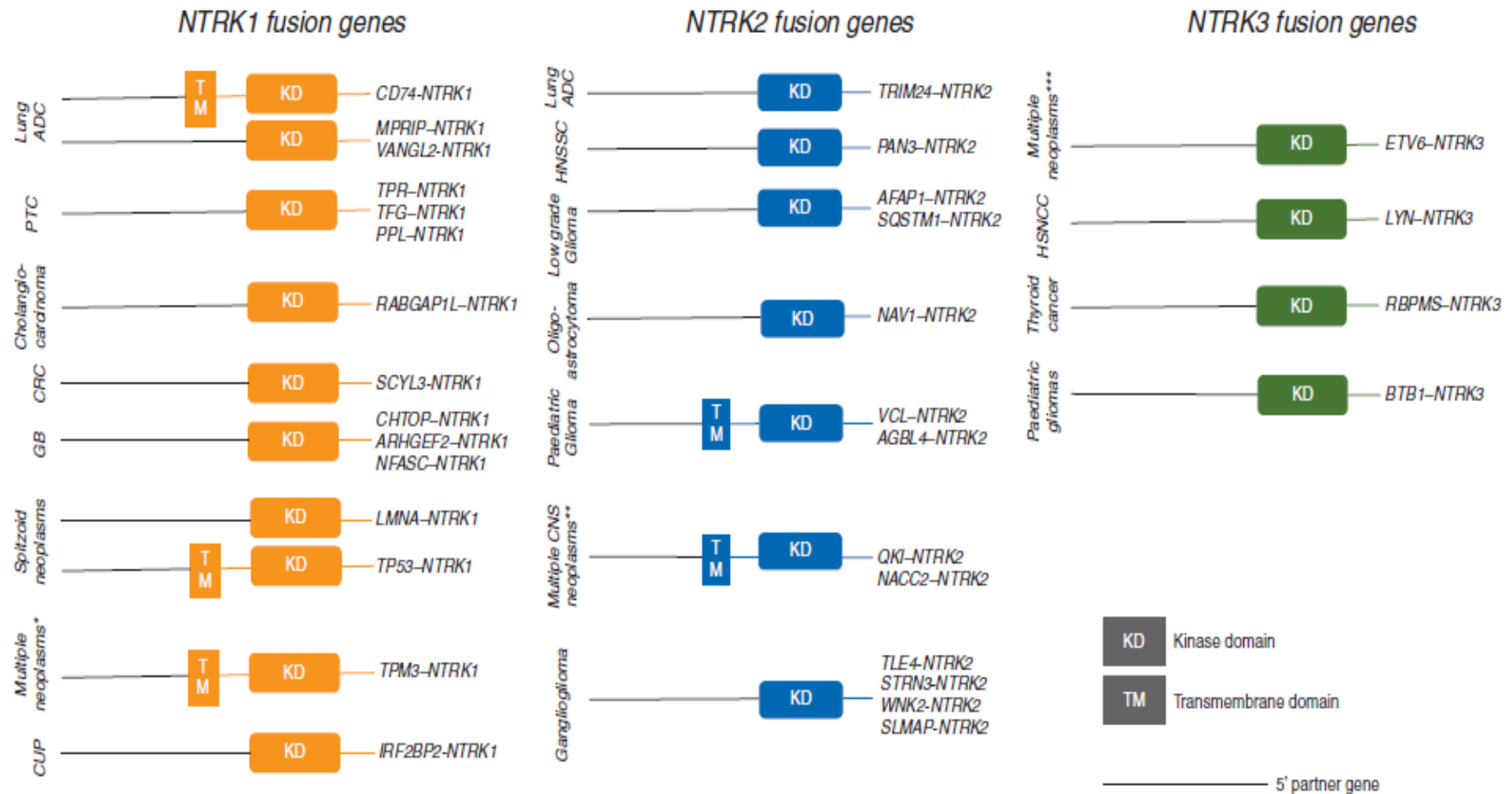
Secretory breast cancer (92%)^{2*}



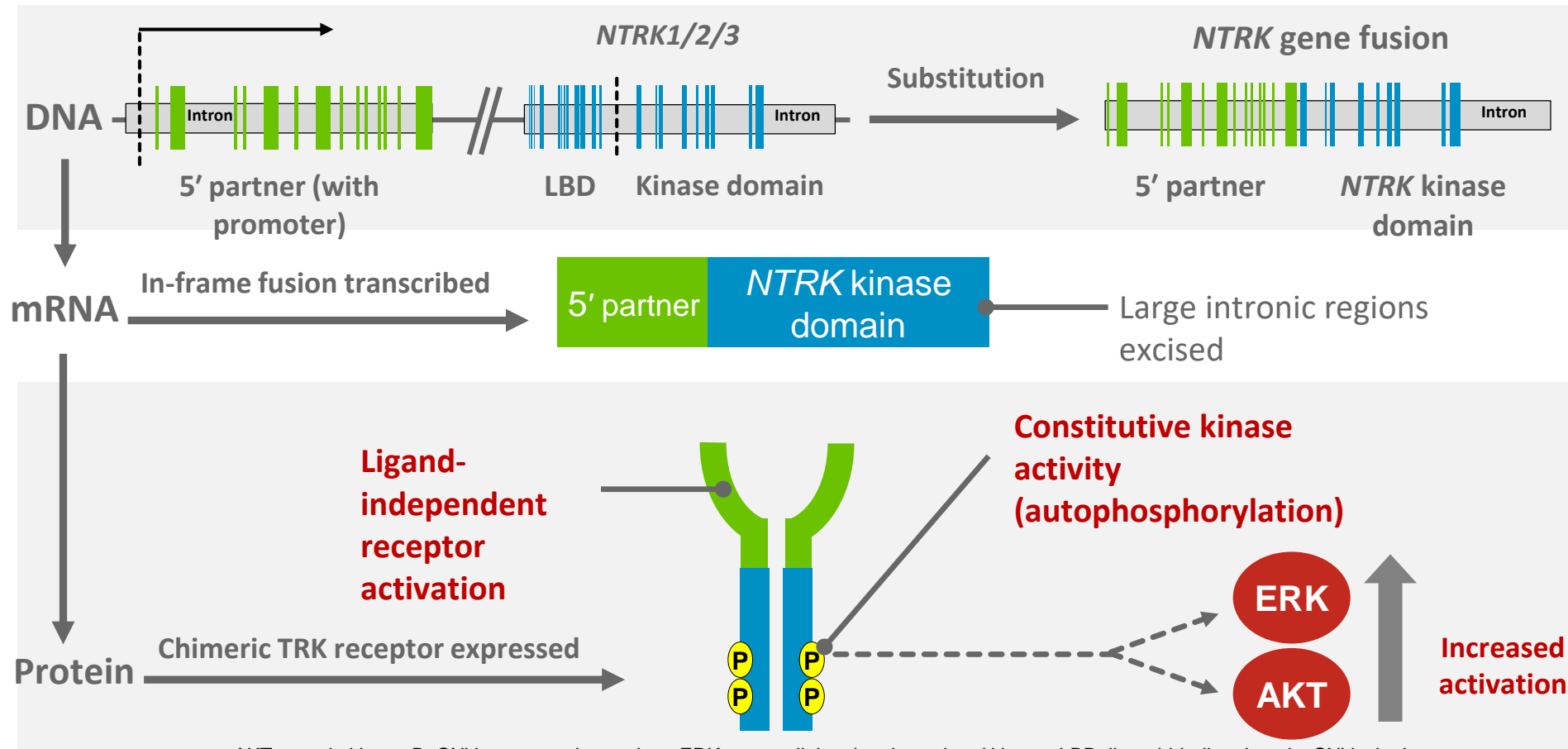
Sarcomas (1%)^{6*}

- *Frequency in adult vs. pediatric patients not specified;
GIST, gastrointestinal stromal tumor;
MASC, mammary analogue secretory carcinoma.

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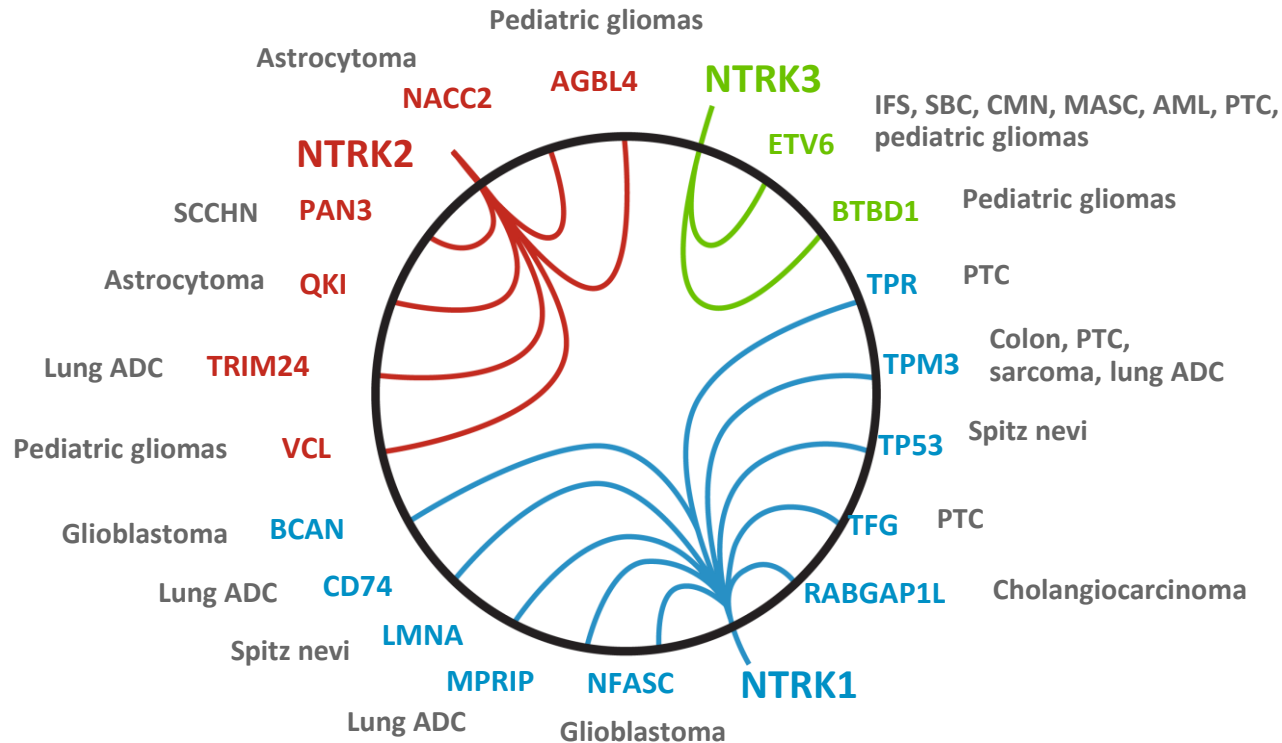
***NTRK* gene fusions are oncogenic – NOT SNVs, CNVs, alterations**



AKT, protein kinase B; CNV, copy number variant; ERK, extracellular signal-regulated kinase; LBD, ligand-binding domain; SNV, single nucleotide variant.

1. Amatu A, et al. *ESMO Open*. 2016;1:e000023.
2. Vaishnavi A, et al. *Cancer Discov*. 2015;5:25-34.
3. Hyman DM, et al. *J Clin Oncol*. 2017;35:LBA2501.

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- *NTRK* gene fusions occur in a tumor-agnostic manner, with inconsistent break points and fusion

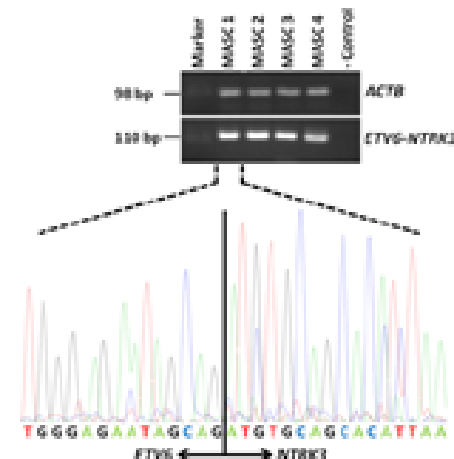
The optimal detection method requires no prior knowledge of fusion break points and/or fusion partner

prehensive *NTRK*

The Art of Identifying *NTRK* Gene Fusions Why Not RT-PCR?

- + High sensitivity and specificity^{1,2}
- + Widely available, low cost³
- Specific primer sets required for each fusion – unknown fusions not detected³⁻⁵
- Low-complexity, GC-rich sequences in *NTRK* genes limit useful primer selection and complicate reaction optimization⁶

Detection of *ETV6-NTRK3* fusion transcripts in MASC tumors by RT-PCR analysis⁷



The Art of Identifying *NTRK* Gene Fusions Why Not FISH?

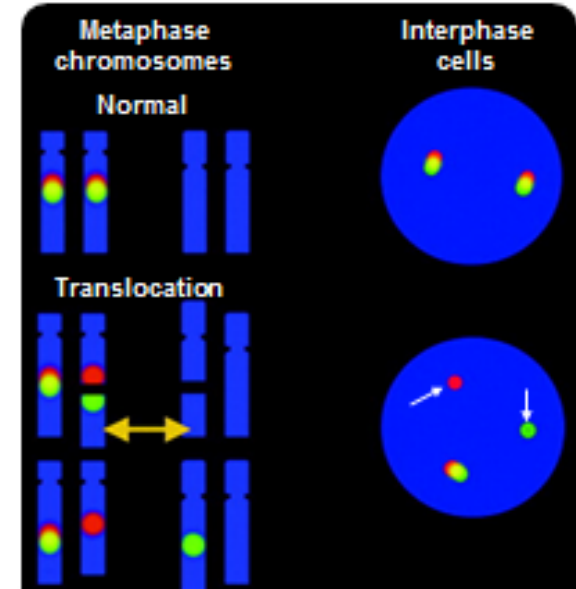
+ Novel fusion partners can be detected if break-apart probes used¹

+ High sensitivity and specificity²

— Requires individual probes for each *NTRK* gene (ie, three separate FISH analyses per patient sample)

— Break-apart FISH identifies gene disruptions in DNA but cannot confirm in-frame, functional fusions

Yellow (red/green) signals in normal interphase nuclei versus split red and green signals in cell carrying translocation



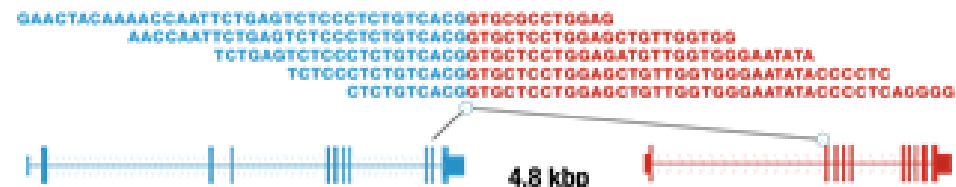
The Art of Identifying *NTRK* Gene Fusions What Makes NGS Ideal?

General NGS

- + High sensitivity and specificity potential
- + Multiplexing: simultaneously queries multiple potentially actionable targets (eg, *NTRK*, *ALK*, *ROS1*, *RET*)
- + Detects both known and novel fusions, regardless of break point or fusion partner (depending on library preparation method)

RNA-based NGS

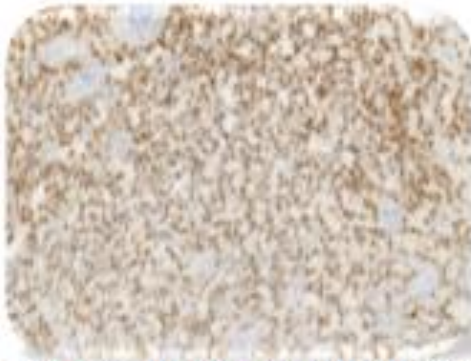
- + Able to distinguish in-frame, transcribed gene fusions versus out-of-frame fusions
- + Avoids difficulties of sequencing large intronic regions in the *NTRK* genes



RNA-based NGS is the preferred method for detecting *NTRK* gene fusions in cancer

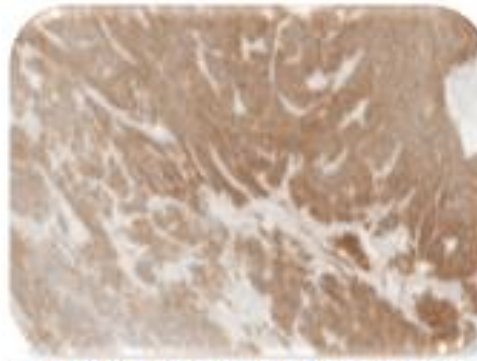
Pan-TRK IHC Detects Both Wild Type and Fusion TRK Protein

Fusion-driven TRK expression



Secretory carcinoma of salivary gland
95% of tumor cells staining
ETV6-NTRK3 fusion detected by NGS

Wild type TRK expression



Gastrointestinal stromal tumor (GIST)
100% tumor cells staining
No fusion detected by NGS

- 9% (n=324) of 3,574 tumors screened by pan-TRK IHC stained positive when defined as >1% of tumor cell staining (range: 0%-54% in gastric cancers and salivary gland cancers, respectively)
- Of the 324 IHC-positive tumors, 164 were confirmed by ISH and 12 fusions were detected
- The percentage of cases staining positive decreased to 5% (n=191) and 4% (n=133) if the definition of positive IHC was increased to >10% and >25% of cells staining, respectively
- At a cutoff of 25% of cells staining, two fusion-positive melanomas would have been missed

Confirmation of IHC-positive cases is REQUIRED

ISH, in situ hybridization
Feng J. et al. Poster presented at: ESMO Molecular Analysis for Personalized Therapy, 2018, Paris, France.

Diagnostic testing methods summary: Benefits and drawbacks

Method				
	Benefits		Drawbacks	
	Next-generation sequencing (NGS)	Fluorescence <i>in situ</i> hybridization (FISH)	Polymerase chain reaction (PCR)	Pan-TRK immunohistochemistry (IHC)
	<ul style="list-style-type: none"> // Detection of novel fusion partners (depending on method)¹ // Ability to interrogate multiple actionable targets simultaneously¹ // Current <i>NTRK</i> testing conducted by NGS² // Relevance of NGS increases as number of actionable targets grows // High sensitivity and specificity potential³ 	<ul style="list-style-type: none"> // Location of the target within the cell is visible^{8,9} // High sensitivity and specificity⁹ // Several fluorophores can be used at once to detect several targets in one sample⁹ 	<ul style="list-style-type: none"> // High sensitivity and specificity^{11,12} // RT-PCR assays detect fusions expressed at the RNA level (also applies to RNA-based NGS)¹² // Inexpensive¹³ 	<ul style="list-style-type: none"> // Inexpensive^{3,14} // Decentralized, available in most laboratories¹¹ // Established reimbursement codes¹⁵ // Turnaround time ~2 days¹⁴
	<ul style="list-style-type: none"> // Turnaround time: ~1-3 weeks // Technically complex (high start-up cost)⁴ // Requires specialty infrastructure⁵ // Highly centralized testing model (academia and reference laboratories)¹ // Reimbursement currently restricted (although developing)⁶ // Sensitivity and specificity of NGS assays vary widely^{3,7} 	<ul style="list-style-type: none"> // Requires fluorescence microscopy¹⁰ // Target sequence must be known (break-apart FISH may detect <i>NTRK</i> gene fusions with unknown partners, but both in-frame and out-of-frame fusions will be detected)¹ 	<ul style="list-style-type: none"> // Target sequences must be known (ie, cannot detect novel fusion partners)^{1,13} 	<ul style="list-style-type: none"> // Detects both fusion and wild-type TRK expression¹⁶ // Scoring algorithms are not standardized¹¹

ESMO recommendations on the standard methods to detect *NTRK* fusions in daily practice and clinical research

Annals of Oncology 30: 1417–1427, 2019
doi:10.1093/annonc/mdz204
Published online 3 July 2019

C. Marchiò^{1,2}, M. Scaltriti^{3,4}, M. Ladanyi³, A. J. Iafrate^{5,6}, F. Bibeau⁷, M. Dietel⁸, J. F. Hechtman³, T. Troiani⁹, F. López-Rios¹⁰, J.-Y. Douillard¹¹, F. André^{12*} & J. S. Reis-Filho³

Table 1. Summary of main features, strengths and weaknesses of all available techniques to detect *NTRK* rearrangements

Method	Sensitivity	Specificity	Detection of all fusion genes	Detection of partner	Detection of expression	Screening
IHC	High ^a	High ^b	Yes	No	Yes	Yes
FISH ^c	High	High	One per probe	No	No	No
RNA seq NGS	High	High	Yes	Yes	Yes	Yes
DNA seq ^c	Moderate	High	Yes	Yes	No	Yes

^aFalse negatives reported mainly in *NTRK3* fusions.

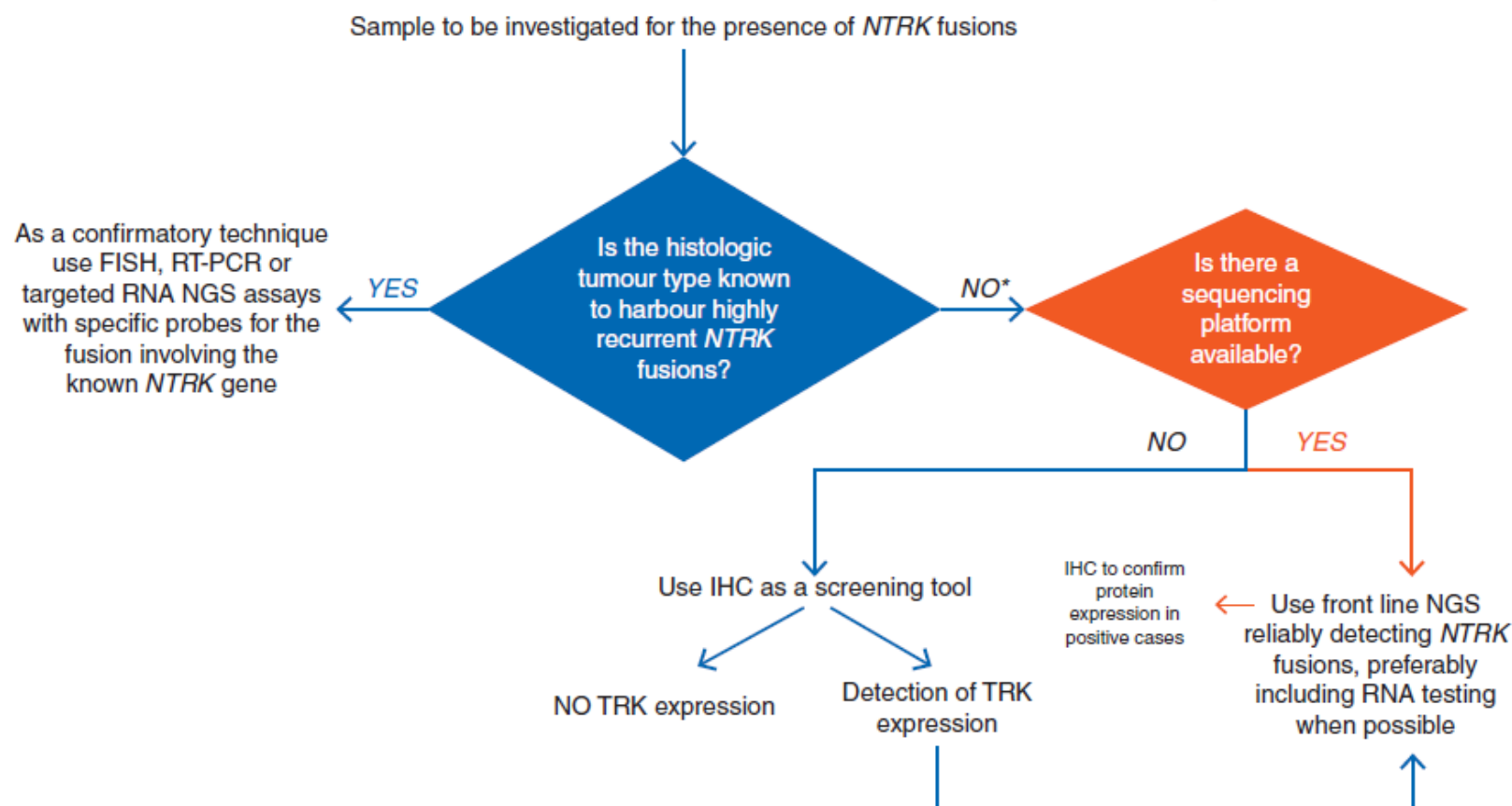
^bIn the absence of smooth muscle/neuronal differentiation.

^cDetected rearrangements by DNA-based assays may not result in fusions, correlation with surgical pathology and predicted transcript (for sequencing) is needed.

ESMO recommendations on the standard methods to detect *NTRK* fusions in daily practice and clinical research

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Testing for *NTRK* gene fusions is essential for identifying patients

NGS, PCR, and FISH can all identify *NTRK* gene fusions

IHC can be used as a screening tool, but results must be confirmed (preferably NGS)

NGS is a multiplex assay able to detect novel fusions and is available at several reference laboratories

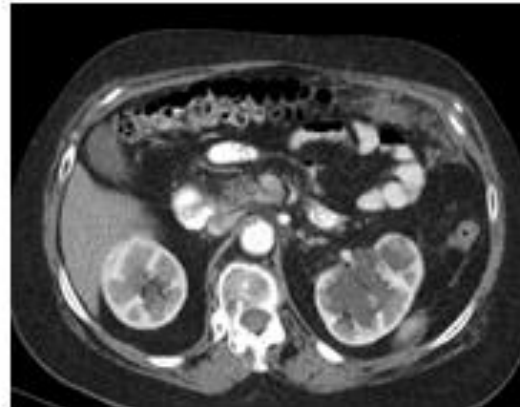
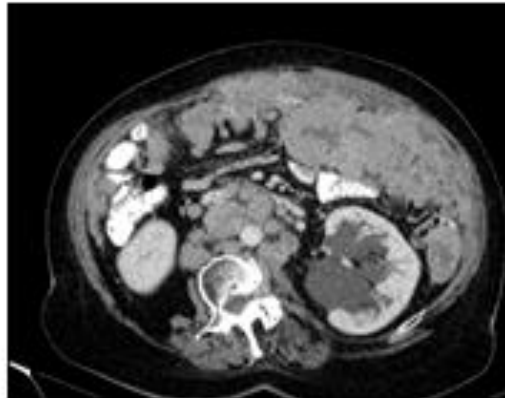
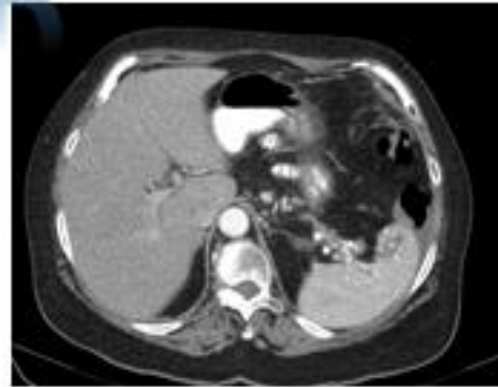
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Oct 2018 CT SCAN



COD

Oct 2019 CT SCAN





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GRACIAS



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